# Package: FieldSimR (via r-universe)

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```
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Maintainer Christian Werner < werner.christian@proton.me>
Description Simulates plot data in multi-environment field trials with
     one or more traits. Its core function generates plot errors
     that capture spatial trend, random error (noise), and
     extraneous variation, which are combined at a user-defined
     ratio. Phenotypes can be generated by combining the plot errors
     with simulated genetic values that capture
     genotype-by-environment (GxE) interaction using wrapper
     functions for the R package `AlphaSimR`.
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Title Simulation of Plot Errors and Phenotypes in Plant Breeding Field

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comps	m_asr_input Simulate genetic values based on a compound symmetry model for	
	GxE interaction - 'AlphaSimR' input parameters	

# Description

Creates a list of input parameters for 'AlphaSimR' to simulate genetic values in multiple environments for one or more traits based on a compound symmetry model for genotype-by-environment (GxE) interaction.

This function utilises the ability of 'AlphaSimR' to simulate correlated traits. The wrapper function compsym\_asr\_input() is used to specify the input parameters required in 'AlphaSimR'. After simulating the genetic values, the wrapper function compsym\_asr\_output can be used to generate a data frame with output values.

# Usage

```
compsym_asr_input(
  ntraits = 1,
  nenvs = 2,
  mean = 0,
```

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```
var = 1,
prop.main = 0.5,
corA = NULL,
meanDD = NULL,
varDD = NULL,
prop.mainDD = NULL,
corDD = NULL,
relAA = NULL,
prop.mainAA = NULL,
corAA = NULL
```

#### Arguments

ntraits Number of traits to be simulated.

nenvs Number of environments to be simulated (minimum of two).

mean A vector of mean genetic values for each environment-within-trait combination.

If only one value is specified, all combinations will be assigned the same mean.

var A vector of genetic variances for each trait.

**Note:** When useVarA = TRUE is specified in 'AlphaSimR' (default), the values in var represent the additive genetic variances, otherwise they represent the total

(additive + non-additive) genetic variances.

prop.main A vector defining the proportion of main effect variance for each trait. If only

one value is specified, all traits will be assigned the same proportion.

**Note:** 0 < prop.main < 1.

corA A matrix of additive genetic correlations between traits. By default, a diagonal

matrix is constructed.

meanDD A vector of mean dominance degrees for each environment-within-trait combi-

nation (similar to mean). If only one value is specified, all combinations will be assigned the same mean. By default, meanDD = NULL and dominance is not

simulated.

varDD A vector of dominance degree variances for each trait.

prop.mainDD A vector defining the proportion of dominance degree main effect variance for

each trait (similar to prop. main). If only one value is specified, all traits will be

assigned the same proportion. **Note:** 0 < prop.mainDD < 1.

corDD A matrix of dominance degree correlations between traits (similar to corA). If

not specified and dominance is simulated, a diagonal matrix is constructed.

relAA A vector defining the relative magnitude of additive-by-additive (epistatic) vari-

ance to additive genetic variance for each trait, that is in a diploid organism with allele frequency of 0.5. If only one value is specified, all traits will be assigned

the same relative magnitude.

prop.mainAA A vector defining the proportion of epistatic main effect variance for each trait

(similar to prop.main). If only one value is specified, all traits will be assigned

the same proportion.

Note: 0 < prop.mainAA < 1.

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corAA

A matrix of epistatic correlations between traits (similar to corA). If not specified and epistasis is simulated, a diagonal matrix is constructed.

#### **Details**

The compound symmetry model assumes the same genetic variance for each environment and the same genetic covariance between each pair of environments. New functionality is being implemented which relaxes the former assumption (also see unstr\_asr\_output).

**Note:** 'AlphaSimR' can simulate different biological effects (see: SimParam).

- For additive traits use addTraitA().
- For additive + dominance traits use addTraitAD().
- For additive + epistatic traits use addTraitAE().
- For additive + dominance + epistatic traits use addTraitADE().

Check the useVarA argument of these functions when simulating non-additive traits.

#### Value

A list with input parameters for 'AlphaSimR', which are used to simulate correlated genetic values based on a compound symmetry model for GxE interaction.

```
# Simulate genetic values with 'AlphaSimR' for two additive + dominance traits
# in two environments based on a compound symmetry model.
# 1. Define the genetic architecture of the simulated traits.
# Mean genetic values and mean dominance degrees.
mean <- c(4.9, 5.4, 235.2, 228.5) # Trait 1 x 2 environments, Trait 2 x 2 environments
meanDD <- c(0.4, 0.4, 0.1, 0.1) # Trait 1 and 2, same value for both environments
# Additive genetic variances and dominance degree variances.
var <- c(0.08, 13) # Different values for Traits 1 and 2
varDD <- 0.2 # Same value for Traits 1 and 2
# Proportion of additive and dominance degree main effect variances.
prop.main <- c(0.4, 0.6) # Different values for Traits 1 and 2
prop.mainDD <- 0.4 # Same value for Traits 1 and 2</pre>
# Additive and dominance degree correlations between the two simulated traits.
corA <- matrix(c(</pre>
 1.0, 0.5,
 0.5, 1.0
), ncol = 2)
corDD <- diag(2) # Assuming independence
input_asr <- compsym_asr_input(</pre>
 ntraits = 2,
 nenvs = 2,
 mean = mean,
```

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```
var = var,
prop.main = prop.main,
corA = corA,
meanDD = meanDD,
varDD = varDD,
prop.mainDD = prop.mainDD,
corDD = corDD
```

compsym\_asr\_output

Simulate genetic values based on a compound symmetry model for GxE interaction - Simulation with 'AlphaSimR'

# **Description**

Creates a data frame of simulated genetic values in multiple environments for one or more traits based on a compound symmetry model for genotype-by-environment (GxE) interaction. The wrapper function compsym\_asr\_output() requires an 'AlphaSimR' population object generated with compsym\_asr\_input.

#### **Usage**

```
compsym_asr_output(pop, ntraits = 1, nenvs, nreps = 1, return.effects = FALSE)
```

# **Arguments**

pop	An 'AlphaSimR' population object (Pop-class or HybridPop-class) generated
	with compsym_asr_input.

ntraits Number of traits specified in compsym\_asr\_input.

nenvs Number of environments specified in compsym\_asr\_input.

nreps A vector defining the number of replicates in each environment. If only one

value is specified, all environments will be assigned the same number.

return.effects When TRUE (default is FALSE), a list is returned with additional entries contain-

ing the genotype main effects and GxE interaction effects for each trait.

#### Value

A data frame with columns 'env', genotype 'id', and 'rep', followed by the simulated genetic values for each trait. When return.effects = TRUE, a list is returned with additional entries containing the genotype main effects and GxE interaction effects for each trait.

```
# Simulate genetic values with 'AlphaSimR' for two additive + dominance traits
# in two environments based on a compound symmetry model.
# 1. Define the genetic architecture of the simulated traits.
# Mean genetic values and mean dominance degrees.
mean <- c(4.9, 5.4, 235.2, 228.5) # Trait 1 x 2 environments, Trait 2 x 2 environments
meanDD <- c(0.4, 0.4, 0.1, 0.1) # Trait 1 and 2, same value for both environments
# Additive genetic variances and dominance degree variances.
var <- c(0.08, 13) # Different values for Traits 1 and 2
varDD <- 0.2 # Same value for Traits 1 and 2
# Proportion of additive and dominance degree main effect variances.
prop.main <- c(0.4, 0.6) # Different values for Traits 1 and 2
prop.mainDD <- 0.4 # Same value for Traits 1 and 2</pre>
# Additive and dominance degree correlations between the two simulated traits.
corA <- matrix(c(</pre>
  1.0, 0.5,
  0.5, 1.0
), ncol = 2)
corDD <- diag(2) # Assuming independence</pre>
input_asr <- compsym_asr_input(</pre>
  ntraits = 2,
 nenvs = 2,
 mean = mean,
  var = var,
  prop.main = prop.main,
  corA = corA,
  meanDD = meanDD,
  varDD = varDD,
  prop.mainDD = prop.mainDD,
  corDD = corDD
)
# 2. Use input_asr to simulate genetic values with 'AlphaSimR' based on a
# compound symmetry model.
library("AlphaSimR")
FOUNDERPOP <- quickHaplo(
  nInd = 10,
  nChr = 1,
  segSites = 20
SP <- SimParam$new(FOUNDERPOP)</pre>
```

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```
SP$addTraitAD(
 nQtlPerChr = 20,
 mean = input_asr$mean,
 var = input_asr$var,
 corA = input_asr$corA,
 meanDD = input_asr$meanDD,
 varDD = input_asr$varDD,
 corDD = input_asr$corDD,
 useVarA = TRUE
# By default, the variances in 'var' represent additive genetic variances.
# When useVarA = FALSE, the values represent total genetic variances.
pop <- newPop(FOUNDERPOP)</pre>
# 3. Create a data frame with simulated genetic values for the two traits in
# the two environments, with two replicates of each genotype.
gv_ls <- compsym_asr_output(</pre>
 pop = pop,
 ntraits = 2,
 nenvs = 2,
 nreps = 2,
 return.effects = TRUE
```

error\_df\_bivar

Plot errors - Example data frame

# **Description**

An example data frame with simulated plot errors for two traits in three environments. Environments 1 and 2 comprise two blocks, while Environment 3 comprises three blocks. The blocks are aligned in the column direction (side-by-side) and comprise 5 columns and 20 rows. The data frame was generated using the function field\_trial\_error with bivariate interpolation. The simulation is demonstrated in the vignette Simulation of plot errors and phenotypes in a plant breeding field trial.

### Usage

```
error_df_bivar
```

### **Format**

A data frame with 700 rows and 6 columns:

env Environment number

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```
block Block numbercol Column numberrow Row numbere.Trait1 Simulated plot errors for Trait 1e.Trait2 Simulated plot errors for Trait 2
```

field\_trial\_error

Simulate plot errors in plant breeding field trials

### **Description**

Creates a data frame of simulated plot errors in multi-environment field trials for one or more traits. The plot errors capture spatial trend, random error (noise), and extraneous variation. Spatial trend is simulated using bivariate interpolation or a separable first-order autoregressive (AR1) process. Random error is simulated using an independent process. Extraneous variation is simulated using random or zig-zag ordering between neighbouring columns and/or rows. The three error components are combined at a user-defined ratio.

Correlated plot errors can be simulated between traits by setting different correlation structures for each error component. A separable structure is assumed between traits and plots within environments, but different error variances can be specified for each environment-within-trait combination.

### Usage

```
field_trial_error(
  ntraits = 1,
  nenvs = 1,
 nblocks = 2,
 block.dir = "col",
  ncols = 10,
  nrows = 20.
  varR = 1,
  ScorR = NULL,
 RcorR = NULL,
 EcorR = NULL,
  spatial.model = "Bivariate",
  complexity = NULL,
  plot.length = 8,
  plot.width = 2,
  col.cor = 0.5,
  row.cor = 0.7,
  prop.spatial = 0.5,
  ext.ord = "random",
  ext.dir = "row",
 prop.ext = 0,
  return.effects = FALSE
)
```

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#### **Arguments**

ntraits Number of traits to be simulated. nenvs Number of environments to be simulated. nblocks A vector defining the number of blocks in each environment. If only one value is specified, all environments will be assigned the same number. block.dir A vector defining the block direction in each environment. Use 'col' for side-byside (default), 'row' for above-and-below, or NA if only one block is simulated. If only one value is specified, all environments will be assigned the same block direction. ncols A vector defining the number of columns in each environment. If only one value is specified, all environments will be assigned the same number. A vector defining the number of rows in each environment. If only one value is nrows specified, all environments will be assigned the same number. A vector of error variances for each environment-within-trait combination. If varR only one value is specified, all combinations will be assigned the same error variance. A matrix of spatial error correlations between traits. If not specified and spatial ScorR trend is simulated, a diagonal matrix is constructed. RcorR A matrix of random error correlations between traits. If not specified and random error is simulated, a diagonal matrix is constructed. EcorR A matrix of extraneous error correlations between traits. If not specified and extraneous variation is simulated, a diagonal matrix is constructed. **Note:** the same correlation between traits is used for the column and row errors. Currently only implemented when ext.ord = "random". spatial.model A character string defining the model used to simulate spatial trend. Use 'Bivariate' for bivariate interpolation (default) or 'AR1' for a separable first-order autoregressive process. Bivariate interpolation is implemented with the interp function of the R package 'interp'. complexity A vector defining the complexity of the simulated spatial trend in each environment when spatial.model = "Bivariate". If only one value is specified, all environments will be assigned the same complexity. If not specified and spatial.model = "Bivariate", the complexity is set to half the maximum number of columns and rows in each environment. plot.length A vector of plot lengths for each environment (column direction). If only one value is specified, all environments will be assigned the same plot length. Only required when spatial.model = "Bivariate". plot.width A vector of plot widths for each environment (row direction). If only one value is specified, all environments will be assigned the same plot width. Only required when spatial.model = "Bivariate". col.cor A vector of column autocorrelations for each environment. If only one value is specified, all environments will be assigned the same column autocorrelation. Only required when spatial.model = "AR1".

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row.cor	A vector of row autocorrelations for each environment. If only one value is specified, all environments will be assigned the same row autocorrelation. Only required when spatial.model = "AR1".
prop.spatial	A vector defining the proportion of spatial trend for each environment-within-trait combination. If only one value is specified, all combinations will be assigned the proportion.
ext.ord	A character string defining the method used to simulate extraneous variation. Use 'random' (default) for random variation between neighbouring columns and/or rows or 'zig-zag' for alternating positive and negative values.
ext.dir	A vector defining the direction of extraneous variation for each environment. Use 'row' (default) for row variation, 'col' for column variation, 'both' for variation in both directions, or NA if no extraneous variation is simulated. When ext.dir = "both", half the variance is assigned to the columns and half is assigned to the rows. If only one value is specified, all environments will be assigned the same direction.
prop.ext	A vector defining the proportion of extraneous variation for each environment-within-trait combination. If only one value is specified, all combinations will be assigned the same proportion.
return.effects	When TRUE (default is FALSE), a list is returned with additional entries containing the spatial, random, and extraneous error terms for each trait.

### Value

A data frame with columns 'env', 'block', 'col', and 'row', followed by the simulated plot errors for each trait. When return.effects = TRUE, a list is returned with additional entries containing the spatial, random, and extraneous error terms for each trait.

```
# Simulate plot errors for two traits in two environments using an AR1 model
# for spatial variation.
# Error variances for the four environment-within-trait combinations.
varR <- c(0.2, 0.4, 10, 15) # Trait 1 x 2 environments, Trait 2 x 2 environments
# Spatial error correlations between the two simulated traits.
ScorR <- matrix(c(</pre>
  1.0, 0.2,
 0.2, 1.0
), ncol = 2)
error_ls <- field_trial_error(</pre>
  ntraits = 2,
  nenvs = 2,
  nblocks = 2,
  block.dir = "row",
  ncols = 10,
  nrows = 20,
  varR = varR,
```

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```
ScorR = ScorR,
spatial.model = "AR1",
col.cor = 0.5,
row.cor = 0.7,
prop.spatial = 0.4,
ext.ord = "zig-zag",
ext.dir = "row",
prop.ext = 0.2,
return.effects = TRUE
)
```

group\_cor\_mat

Simulate a reduced rank correlation matrix with multiple groups

# Description

Creates a symmetric correlation matrix with user-defined structure, rank and groupings.

# Usage

```
group_cor_mat(
  n = c(5, 5),
  within.cor = 0.5,
  between.cor = 0.2,
  range = NULL,
  rank = 4,
  skew = 0,
  pos.def = FALSE,
  small.positive = NULL,
  return.groups = FALSE
)
```

# **Arguments**

n	A vector defining the size of each group.
within.cor	A vector defining the baseline correlation within each group. If only one value is supplied, all groups will be assigned the same correlation.  Note: -1 < within.cor < 1.
between.cor	A scalar defining the baseline correlation between groups.  Note: between.cor <= within.cor.
range	A scalar defining the range of correlations around the baseline. By default, range = 1 - max(within.cor) which ensures the matrix is positive semi-definite.  Note: max(within.cor) + range <= 1.
rank	A scalar defining the rank of the correlation matrix.
skew	A scalar defining the skewness imposed on the correlations. <b>Note:</b> $-1 < \text{skew} < 1$ .

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When TRUE (default is FALSE), the function bend of the R package 'mbend' is used to bend a non-positive (semi)-definite matrix to be positive (semi)-definite.

small.positive Argument passed to bend when pos.def = TRUE (default is 1e-8). Eigenvalues smaller than small.positive are replaced by this.

Note: 0 < small.positive < 0.1.

return.groups When TRUE (default is FALSE), a list is returned with additional entries containing the members of each group.

#### Value

A symmetric correlation matrix with defined rank and groupings. When pos.def = TRUE, the correlation matrix is guaranteed to be positive (semi)-definite. When return.groups = TRUE, a list is returned with additional entries containing the group members.

# **Examples**

```
# Simulate and visualise a correlation matrix with 2 groups containing 5 and 10 members,
# correlations of 0.4 within groups and 0 between groups and rank equal to 4

cor_ls <- group_cor_mat(
    n = c(5, 10),
    within.cor = 0.4,
    between.cor = 0,
    rank = 4,
    return.groups = TRUE
)

plot_matrix(
    mat = cor_ls$cor.mat,
    group.df = cor_ls$group.df,
    order = TRUE
)</pre>
```

gv\_df\_unstr

Genetic values - Example data frame

# **Description**

An example data frame with simulated genetic values of 100 genotypes for two traits in three environments. Environments 1 and 2 comprise two replicates of each genotype, while Environment 3 comprises three replicates. The data frame was generated using the wrapper functions unstr\_asr\_input and unstr\_asr\_output, which simulate correlated genetic values with 'AlphaSimR'. The simulation is demonstrated in the vignette Simulation of genetic values based on an unstructured model for GxE interaction.

### Usage

```
gv_df_unstr
```

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# **Format**

```
A data frame with 700 rows and 5 columns:
```

```
env Environment number
id Genotype identifier
rep Replicate number
gv.Trait1 Simulated genetic values for Trait 1
gv.Trait2 Simulated genetic values for Trait 2
```

make\_phenotypes

Generate phenotypes - Combine genetic values and plot errors

# **Description**

Creates a data frame of phenotypes by combining genetic values with plot errors generated with the function field\_trial\_error. Requires genetic values generated with the functions compsym\_asr\_output or unstr\_asr\_output, or any data frame matching the description below.

# Usage

```
make_phenotypes(
  gv.df,
  error.df,
  design.df = NULL,
  randomise = TRUE,
  return.effects = FALSE
)
```

# Arguments

gv.df	A data frame of genetic values. Must contain the columns 'env', genotype 'id', 'rep', and the genetic values for each trait.
error.df	A data frame of plot errors. Must contain the columns 'env', 'block', 'col', 'row', and the plot errors for each trait.
design.df	A optional data frame of frequencies for generating incomplete block designs. Must contain the columns 'env', 'id', and 'nreps' indicating the number of replicates per individual for each environment.
randomise	When TRUE (default), genotypes are randomly allocated to plots according to a randomized complete (or incomplete) block design.  Note: Other experimental designs are being implemented and should be generated externally.
return.effects	When TRUE (default is FALSE), a list is returned with additional entries containing the genetic values and plot errors for each trait.

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#### Value

A data frame with columns 'env', 'block', 'column', 'row', genotype 'id', 'rep', and the phenotypes for each trait. When return.effects = TRUE, a list is returned with additional entries containing the genetic values and plot errors for each trait.

# **Examples**

# Generate and visualise phenotypes by combining the genetic values and plot errors provided # in the two example data frames  $gv_df_unstr$  and  $error_df_bivar$ .

```
pheno_ls <- make_phenotypes(
   gv.df = gv_df_unstr,
   error.df = error_df_bivar,
   randomise = TRUE,
   return.effects = TRUE
)

plot_effects(
   df = pheno_ls$pheno.df[pheno_ls$pheno.df$env == 1, ],
   effect = "y.Trait1",
   labels = TRUE,
)</pre>
```

measure\_variances

Measure variances from a covariance matrix

# **Description**

Creates a table of variance components derived from a user-defined covariance matrix.

### Usage

```
measure_variances(mat, correction = FALSE)
```

### **Arguments**

mat A symmetric n x n positive (semi)-definite variance matrix.

correction When TRUE (default is FALSE), the variance components are treated as estimates

and sample corrections are applied.

### Value

A table which partitions the total variance into main effect and interaction variance, heterogeneity of variance and lack of correlation, and non-crossover and crossover interaction. When correction = TRUE, a sample correction is applied to the variance components.

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### **Examples**

# Generate a structured covariance matrix and then measure the partitioning of variance.

```
diag_mat <- rand_diag_mat(
    n = 10
)
cor_mat <- struc_cor_mat(
    n = 10,
)

cov_mat <- sqrt(diag_mat) %*% cor_mat %*% sqrt(diag_mat)
measure_variances(
    mat = cov_mat
)</pre>
```

multi\_asr\_input

Simulate genetic values based on a multiplicative model for GxE interaction - 'AlphaSimR' input parameters

# **Description**

Creates a list of input parameters for 'AlphaSimR' to simulate genetic values in multiple environments for one or more traits based on a (reduced rank) multiplicative model for genotype-by-environment (GxE) interaction.

This function utilises the ability of 'AlphaSimR' to simulate correlated traits. The wrapper function multi\_asr\_input() is used to specify the input parameters required in 'AlphaSimR'. After simulating the genetic values, the wrapper function multi\_asr\_output can be used to generate a data frame with output values.

### Usage

```
multi_asr_input(
  ntraits = 1,
  nenvs = 2,
  mean = 0,
  var = 1,
  corA = NULL,
  nterms = NULL
)
```

### **Arguments**

ntraits Number of traits to be simulated.

nenvs Number of environments to be simulated (minimum of two).

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mean	A vector of mean genetic values for each trait or each environment-within-trait combination. If only one value is specified, all combinations will be assigned the same mean.
var	A vector of additive genetic variances for each trait or each environment-within- trait combination. If only one value is specified, all combinations will be as- signed the same variance.
corA	A matrix of additive genetic correlations between environment-within-trait combinations. By default, a diagonal matrix is constructed.
nterms	A scalar defining the number of multiplicative terms to be simulated. By default, the number of terms is set to the number of environment-within-trait combinations. <b>Note:</b> when nterms is less than the number of environment-within-trait combinations, the values in mean will be approximated.

#### **Details**

Currently supports additive traits only, but other (non-additive) traits are being implemented.

#### Value

A list with input parameters for 'AlphaSimR', which are used to simulate correlated genetic values based on a multiplicative model for GxE interaction. Covariates are also supplied for use in multi\_asr\_output.

```
# Simulate genetic values with 'AlphaSimR' for two additive traits in two
# environments based on a multiplicative model with three terms.
# 1. Define the genetic architecture of the simulated traits.
# Mean genetic values.
mean <- c(5, 240) # Trait 1, Trait 2
# Additive genetic variances.
var <- c(0.086, 0.12, 15.1, 8.5) # Trait 1 x 2 environments, Trait 2 x 2 environments
# Additive genetic correlations between the two simulated traits.
TcorA <- matrix(c(</pre>
  1.0, 0.6,
  0.6, 1.0
), ncol = 2)
# Additive genetic correlations between the two simulated environments.
EcorA <- matrix(c(</pre>
  1.0, 0.2,
  0.2, 1.0
), ncol = 2)
# Construct separable additive genetic correlation matrix.
corA <- kronecker(TcorA, EcorA)</pre>
```

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```
input_asr <- multi_asr_input(
  ntraits = 2,
  nenvs = 2,
  mean = mean,
  var = var,
  corA = corA,
  nterms = 3
)</pre>
```

multi\_asr\_output

Simulate genetic values based on a multiplicative model for GxE interaction - Simulation with 'AlphaSimR'

# Description

Creates a data frame of simulated genetic values in multiple environments for one or more traits based on a (reduced rank) multiplicative model for genotype-by-environment (GxE) interaction. This function requires an 'AlphaSimR' population object generated with multi\_asr\_input.

# Usage

```
multi_asr_output(
  pop,
  ntraits = 1,
  nenvs,
  nreps = 1,
  cov.mat,
  return.effects = FALSE
)
```

# Arguments

pop	An 'AlphaSimR' population object (Pop-class or HybridPop-class) generated with multi_asr_input.
ntraits	Number of traits specified in multi_asr_input.
nenvs	Number of environments specified in multi_asr_input.
nreps	A vector defining the number of replicates in each environment. If only one value is specified, all environments will be assigned the same number.
cov.mat	A matrix of covariates that will be used to construct the genetic values, typically generated with multi_asr_input.
return.effects	When TRUE (default is FALSE), a list is returned with additional entries containing the genotype slopes for each multiplicative term.

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#### Value

A data frame with columns 'env', genotype 'id', and 'rep', followed by the simulated genetic values for each trait. When return.effects = TRUE, a list is returned with additional entries containing the genotype slopes for each multiplicative term.

```
# Simulate genetic values with 'AlphaSimR' for two additive traits in two
# environments based on a multiplicative model with three terms.
# 1. Define the genetic architecture of the simulated traits.
# Mean genetic values.
mean <- c(5, 240) # Trait 1, Trait 2
# Additive genetic variances.
var <- c(0.086, 0.12, 15.1, 8.5) # Trait 1 x 2 environments, Trait 2 x 2 environments
# Additive genetic correlations between the two simulated traits.
TcorA <- matrix(c(</pre>
  1.0, 0.6,
  0.6, 1.0
), ncol = 2)
# Additive genetic correlations between the two simulated environments.
EcorA <- matrix(c(</pre>
 1.0, 0.2,
  0.2, 1.0
), ncol = 2)
# Construct separable additive genetic correlation matrix
corA <- kronecker(TcorA, EcorA)</pre>
input_asr <- multi_asr_input(</pre>
  ntraits = 2,
  nenvs = 2,
 mean = mean,
 var = var,
  corA = corA,
  nterms = 3
)
# 2. Use input_asr to simulate genetic values in 'AlphaSimR' based on a
# multiplicative model with three terms.
library("AlphaSimR")
FOUNDERPOP <- quickHaplo(
  nInd = 10,
  nChr = 1,
  segSites = 20
)
```

plot\_effects 19

```
SP <- SimParam$new(FOUNDERPOP)</pre>
SP$addTraitA(
  nQtlPerChr = 20,
  mean = input_asr$mean,
  var = input_asr$var,
  corA = input_asr$corA
)
pop <- newPop(FOUNDERPOP)</pre>
# 3. Create a data frame with simulated genetic values for the two traits in the two
# environments, with two replicates of each genotype.
# The covariates are obtained from input_asr.
gv_ls <- multi_asr_output(</pre>
  pop = pop,
 ntraits = 2,
 nenvs = 2,
  nreps = 2,
  cov.mat = input_asr$cov.mat,
  return.effects = TRUE
```

plot\_effects

Graphics for plot effects

# **Description**

Creates a graphical field array for a set of plot effects (e.g., phenotypes, genetic values, or plot errors). Requires a data frame generated with the functions field\_trial\_error or make\_phenotypes, or any data frame matching the description below.

# Usage

```
plot_effects(df, effect, blocks = TRUE, labels = TRUE)
```

# **Arguments**

at	A data frame with the columns 'col', 'row', and the effects to be plotted.
effect	The name of the effects to be plotted.
blocks	When TRUE (default), the field array is split into blocks. This requires an additional column 'block' in the data frame.
labels	When TRUE (default), column and row labels are displayed.

20 plot\_hist

### Value

A graphical field array with x- and y-axes displaying the column and row numbers, and colour gradient ranging from red (low value) to green (high value).

### **Examples**

```
# Display the simulated plot errors in the example data frame 'error_df_bivar'
# for Trait 1 in Environment 1.
error_df <- error_df_bivar[error_df_bivar$env == 1, ]

plot_effects(
    df = error_df,
    effect = "e.Trait1",
    labels = TRUE,
)</pre>
```

plot\_hist

Histogram of values

### **Description**

Creates a histogram of user-defined values (e.g., effects, correlations, or covariances).

# Usage

```
plot_hist(df, value = NULL, bins = 30, density = FALSE)
```

# **Arguments**

A data frame or vector with the values to be plotted.

The name of the values to be plotted. Ignored when 'df' is a vector.

Argument passed to ggplot2 (default is 30). Controls the number of bins in the histogram.

When TRUE (default is FALSE), a density curve is superimposed onto the his-

togram.

#### Value

A histogram with x- and y-axes displaying the values and their frequency, respectively. When density = TRUE, a density curve is superimposed onto the histogram.

plot\_matrix 21

### **Examples**

```
# Histogram of the simulated plot errors in the example data frame 'error_df_bivar'
# for Trait 1 in Environment 1.
error_df <- error_df_bivar[error_df_bivar$env == 1, ]
plot_hist(
    df = error_df,
    value = "e.Trait1",
    density = TRUE
)</pre>
```

plot\_matrix

Graphics for matrices

# Description

Creates a heatmap of a symmetric matrix (e.g., correlation or covariance matrix).

# Usage

```
plot_matrix(mat, order = FALSE, group.df = NULL, labels = TRUE)
```

# **Arguments**

mat A symmetric matrix.

order When TRUE (default is FALSE), the function agnes of the R package 'cluster' is used with default arguments to order the matrix based on a dendrogram.

group.df An optional data frame with columns containing the variable names followed by the group numbers. When supplied, the heatmap is split into groups and then ordered (when order = TRUE).

labels When TRUE (default), variable labels are displayed.

#### Value

A heatmap with x- and y-axes displaying the variable numbers, and colour gradient ranging from blue (low value) to red (high value).

```
# Display a random correlation matrix.
cor_mat <- rand_cor_mat(
    n = 10,
    min.cor = -1,
    max.cor = 1
)
# Define groups.</pre>
```

22 qq\_plot

```
group_df <- data.frame(variable = 1:10, group = c(1, 1, 1, 1, 2, 2, 2, 3, 3, 4))
plot_matrix(
  mat = cor_mat,
    group.df = group_df,
  order = TRUE,
  labels = TRUE
)</pre>
```

qq\_plot

Q-Q plot

# Description

Creates a normal quantile-quantile (Q-Q) plot for a set of effects (e.g., phenotypes, genetic values, or plot errors).

# Usage

```
qq_plot(df, effect, labels = FALSE)
```

### **Arguments**

df A data frame or vector with the effects to be plotted.

effect The name of the effects to be plotted. Ignored when 'df' is a vector.

labels When TRUE (default is FALSE), column and row labels are displayed. This re-

quires additional columns 'col' and 'row' in the data frame.

#### Value

A Q-Q plot with x- and y-axes displaying the theoretical and sample quantiles of the effects, respectively.

```
# Q-Q plot of the simulated plot errors in the example data frame 'error_df_bivar'
# for Trait 1 in Environment 1.

error_df <- error_df_bivar[error_df_bivar$env == 1, ]

qq <- qq_plot(
    df = error_df,
    effect = "e.Trait1",
    labels = TRUE
)

# Q-Q plot
qq</pre>
```

rand\_cor\_mat 23

```
\# Extract the data frame with the theoretical and sample quantiles of the \# user-defined effects. 
 qq_df <- qq$data
```

rand\_cor\_mat

Simulate a random correlation matrix

# Description

Creates a symmetric n x n correlation matrix with user-defined minimum and maximum correlations based on a continuous uniform distribution.

# Usage

```
rand_cor_mat(
  n = 5,
  min.cor = -1,
  max.cor = 1,
  pos.def = FALSE,
  small.positive = NULL
)
```

# Arguments

n	A scalar defining the dimensions of the correlation matrix.
min.cor	A scalar defining the minimum correlation.
max.cor	A scalar defining the maximum correlation.  Note: -1 < min.cor < max.cor < 1.
pos.def	When TRUE (default is FALSE), the function bend of the R package 'mbend' is used to bend a non-positive (semi)-definite matrix to be positive (semi)-definite.
small.positive	Argument passed to bend when pos.def = TRUE (default is 1e-8). Eigenvalues smaller than small.positive are replaced by this.  Note: $0 < \text{small.positive} < 0.1$ .

### Value

A symmetric  $n \times n$  correlation matrix. When pos. def = TRUE, the correlation matrix is guaranteed to be positive (semi)-definite.

24 rand\_diag\_mat

### **Examples**

```
# Simulate and visualise a random correlation matrix with 10 columns and rows.
cor_mat <- rand_cor_mat(
    n = 10,
    min.cor = -0.2,
    max.cor = 0.8,
    pos.def = TRUE
)

plot_matrix(
    mat = cor_mat,
    order = TRUE
)</pre>
```

rand\_diag\_mat

Simulate a random diagonal variance matrix

# Description

Creates a diagonal n x n variance matrix with user-defined minimum and maximum variances based on a continuous uniform distribution.

### Usage

```
rand_diag_mat(n = 5, min.var = 0, max.var = 1)
```

# **Arguments**

n A scalar defining the dimensions of the variance matrix.

min.var A scalar defining the minimum variance.

max.var A scalar defining the maximum variance.

Note: 0 < min.var < max.var.

# Value

A diagonal n x n variance matrix.

```
# Simulate a random diagonal matrix with 10 columns and rows.
diag_mat <- rand_diag_mat(
    n = 10,
    min.var = 0,
    max.var = 0.2
)</pre>
```

sample\_met 25

sample_met	Sample environments from a target population

# **Description**

Creates a list of environments sampled from a population with user-defined sample size.

# Usage

```
sample_met(
  ntraits = 1,
  nenvs = 1000,
  nsamples = 10,
  sample.size = 20,
  replace = TRUE,
  cov.mat = NULL
)
```

# Arguments

ntraits	A scalar defining the number of traits.
nenvs	A scalar defining the number of environments in the target population.
nsamples	A scalar defining the number of samples to be taken.
sample.size	A vector defining the number of environments in each sample. When only one value is specified, all samples will be assigned the same number.
replace	When TRUE (default), samples are taken with replacement. Ignored when nsamples = 1.
cov.mat	An optional matrix of environmental covariates for one or more traits. When supplied, the covariates are sampled and printed.

# Value

A list with elements given by the sample of environments taken from the target population. When cov.mat is supplied, additional entries are given containing the sampled environmental covariates for each trait.

```
# Sample environments from a target population of 1000, with each sample containing 20 environments.
cov_ls <- sample_met(
  nenvs = 1000,
   nsamples = 10,
  sample.size = 20,
  replace = TRUE
)</pre>
```

26 sample\_variogram

sample\_variogram

Sample variogram

# **Description**

Creates a sample variogram for a set of effects (e.g., plot errors).

### Usage

```
sample_variogram(df, effect, min.np = 30)
```

# Arguments

df A data frame with the columns 'col', 'row', and the effects to be plotted.

effect The name of the effects to be plotted.

min.np Minimum number of pairs for which semivariances are displayed (default is 30).

# Value

A sample variogram with x- and y-axes displaying the row and column displacements, and the z-axis displaying the average semivariances (variogram ordinates) for the effects.

```
# Sample variogram of plot errors simulated using a separable first order
# autoregressive (AR1) process.
error_df <- field_trial_error(</pre>
 ntraits = 1,
 nenvs = 1,
 spatial.model = "AR1"
)
variogram <- sample_variogram(</pre>
 df = error_df,
 effect = "e.Trait1"
)
# Sample variogram
variogram
# Extract the data frame with the column and row displacements, and the
# average semivariances.
variogram_df <- variogram$data</pre>
```

skew\_diag\_mat 27

skew_diag_mat	Simulate a skewed diagonal variance matrix	

# Description

Creates a diagonal n x n variance matrix with user-defined skewness based on a gamma or inverse gamma distribution.

# Usage

```
skew_diag_mat(n = 5, shape = 1.5, scale = 1, inverse = FALSE, mean.var = NULL)
```

# Arguments

n	A scalar defining the dimensions of the variance matrix.
shape	A scalar defining the shape of the distribution.
scale	A scalar defining the scale of the distribution.
inverse	When TRUE (default is FALSE), the variances are sampled from the inverse gamma distribution instead of the gamma distribution.
mean.var	An optional scalar defining the mean variance When supplied, the variances are scaled to achieve the defined mean.

### Value

A diagonal n x n variance matrix.

```
# Simulate a random diagonal matrix with 10 columns and rows, and negatively skewed variances
# scaled to a mean of 0.1.
diag_mat <- skew_diag_mat(
    n = 10,
    shape = 1.5,
    scale = 1,
    mean.var = 0.1
)</pre>
```

28 struc\_cor\_mat

struc_cor_mat	Simulate a structured correlation matrix with reduced rank

# Description

Creates a symmetric n x n correlation matrix with user-defined structure and rank.

# Usage

```
struc_cor_mat(
  n = 5,
  base.cor = 0.5,
  range = NULL,
  rank = 3,
  skew = 0,
  base.mat = NULL,
  pos.def = FALSE,
  small.positive = NULL
)
```

# Arguments

n	A scalar defining the dimensions of the correlation matrix.
base.cor	A scalar defining the baseline correlation.  Note: -1 < base.cor < 1.
range	A scalar defining the range of correlations around the baseline. By default, range = 1 - base.cor which ensures the matrix is positive semi-definite with defined rank. <b>Note:</b> base.cor + range <= 1.
rank	A scalar defining the rank of the correlation matrix.
skew	A scalar defining the skewness imposed on the correlations. Note: $-1 < skew < 1$ .
base.mat	An optional $n \times n$ base correlation matrix. When supplied, base .cor and skew are ignored and noise is simulated based on rank.
pos.def	When TRUE (default is FALSE), the function bend of the R package 'mbend' is used to bend a non-positive (semi)-definite matrix to be positive (semi)-definite.
small.positive	Argument passed to bend when pos.def = TRUE (default is 1e-8). Eigenvalues smaller than small.positive are replaced by this. Note: $0 < \text{small.positive} < 0.1$ .

# Value

A symmetric  $n \times n$  correlation matrix with defined rank. When pos.def = TRUE, the correlation matrix is guaranteed to be positive (semi)-definite.

theoretical\_variogram 29

### **Examples**

```
# Simulate and visualise a correlation matrix with 10 columns and rows, rank equal to 4 and
# negatively skewed correlations.
cor_mat <- struc_cor_mat(
    n = 10,
    base.cor = 0.3,
    range = 0.7,
    rank = 4,
    skew = -0.5
)

plot_matrix(
    mat = cor_mat,
    order = TRUE
)</pre>
```

theoretical\_variogram Theoretical variogram

### **Description**

Creates a theoretical variogram for a separable first order autoregressive (AR1) process.

### Usage

```
theoretical_variogram(
  ncols = 10,
  nrows = 20,
  varR = 1,
  col.cor = 0.5,
  row.cor = 0.7,
  prop.spatial = 1
)
```

# **Arguments**

```
ncols A scalar defining the number of columns.

nrows A scalar defining the number of rows.

varR A scalar defining the error variance.

col.cor A scalar defining the column autocorrelation,

row.cor A scalar defining the row autocorrelation.

prop.spatial A scalar defining the proportion of spatial trend.
```

#### Value

A theoretical variogram with x- and y-axes displaying the row and column displacements, and the z-axis displaying the semivariances (variogram ordinates) for a separable autoregressive process.

### **Examples**

```
# Theoretical variogram for a field trial with 10 columns and 20 rows, based
# on column and row autocorrelations of 0.5 and 0.7, and a proportion of
# spatial trend of 0.5. The remaining proportion represents random error.
variogram <- theoretical_variogram(</pre>
 ncols = 10,
 nrows = 20,
 varR = 1,
 col.cor = 0.5,
 row.cor = 0.7,
 prop.spatial = 0.5
)
# Theoretical variogram
variogram
# Extract the data frame with the column and row displacements, and the
# theoretical semivariances.
variogram_df <- variogram$data</pre>
```

unstr\_asr\_input

Simulate genetic values based on an unstructured model for GxE interaction - 'AlphaSimR' input parameters

### **Description**

Creates a list of input parameters for 'AlphaSimR' to simulate genetic values in multiple environments for one or more traits based on an unstructured model for genotype-by-environment (GxE) interaction.

This function utilises the ability of 'AlphaSimR' to simulate correlated traits. The wrapper function unstr\_asr\_input() is used to specify the input parameters required in 'AlphaSimR', and can handle separable and non-separable structures between traits and environments (see below). After simulating the genetic values, the wrapper function unstr\_asr\_output can be used to generate a data frame with output values.

# Usage

```
unstr_asr_input(
  ntraits = 1,
  nenvs = 2,
  mean = 0,
  var = 1,
  Tvar = NULL,
  Evar = NULL,
  corA = NULL,
  TcorA = NULL,
```

```
EcorA = NULL,
meanDD = NULL,
varDD = NULL,
EvarDD = NULL,
corDD = NULL,
EcorDD = NULL,
EcorDD = NULL,
relAA = NULL,
TrelAA = NULL,
ErelAA = NULL,
CorAA = NULL,
EcorAA = NULL)
```

### **Arguments**

ntraits Number of traits to be simulated.

nenvs Number of environments to be simulated (minimum of two).

mean A vector of mean genetic values for each environment-within-trait combination.

If only one value is specified, all combinations will be assigned the same mean.

var A vector of genetic variances for each environment-within-trait combination. If

only one value is specified, all combinations will be assigned the same variance. **Alternatively**, if a separable structure between traits and environments is de-

sired, Tvar and Evar can be specified.

Tvar A vector of genetic variances for each trait. Must be provided in combination

with Evar.

Alternatively, var can be specified.

Evar A vector of genetic variances for each environment. Must be provided in com-

bination with Tvar.

Alternatively, var can be specified.

corA A matrix of additive genetic correlations between environment-within-trait com-

binations. By default, a diagonal matrix is constructed.

Alternatively, ToorA and EcorA can be specified.

TcorA A matrix of additive genetic correlations between traits. Must be provided in

combination with EcorA.

**Alternatively**, corA can be specified.

EcorA A matrix of additive genetic correlations between environments. Must be pro-

vided in combination with TcorA.

Alternatively, corA can be specified.

meanDD A vector of mean dominance degrees for each environment-within-trait combi-

nation (similar to mean). If only one value is specified, all combinations will be assigned the same mean. By default, meanDD = NULL and dominance is not

simulated.

varDD A vector of dominance degree variances for each environment-within-trait combination (similar to var). If only one value is specified, all combinations will be assigned the same variance.

**Alternatively**, if a separable structure between traits and environments is desired, TvarDD and EvarDD can be specified.

TvarDD A vector of dominance degree variances for each trait (similar to Tvar). Must

be provided in combination with EvarDD. **Alternatively**, varDD can be specified.

EvarDD A vector of dominance degree variances for each environment (similar to Evar).

Must be provided in combination with TvarDD.

Alternatively, varDD can be specified.

corDD A matrix of dominance degree correlations between environment-within-trait

combinations (similar to corA). If not specified and dominance is simulated, a

diagonal matrix is constructed.

**Alternatively**, TcorDD and EcorDD can be specified.

TcorDD A matrix of dominance degree correlations between traits (similar to TcorA).

Must be provided in combination with EcorDD.

**Alternatively**, corDD can be specified.

EcorDD A matrix of dominance degree correlations between environments (similar to

EcorA). Must be provided in combination with TcorDD.

Alternatively, corDD can be specified.

relAA A vector defining the relative magnitude of additive-by-additive (epistatic) vari-

ance to additive genetic variance for each environment-within-trait combination, that is in a diploid organism with allele frequency of 0.5. If only one value is specified, all environment-within-trait combinations will be assigned the same

value. By default, relAA = NULL and epistasis is not simulated.

**Alternatively**, if a separable structure between traits and environments is desired, TrelAA and ErelAA can be specified.

TrelAA A vector defining the relative magnitude of epistatic variance to additive genetic

variance for each trait. Must be provided in combination with ErelAA.

Alternatively, relAA can be specified.

ErelAA A vector defining the relative magnitude of epistatic variance to additive genetic

variance for each environment. Must be provided in combination with TrelAA.

**Alternatively**, relAA can be specified.

corAA A matrix of epistatic correlations between environment-within-trait combina-

tions (similar to corA). If not specified and epistasis is simulated, a diagonal

matrix is constructed.

Alternatively, TcorAA and EcorAA can be specified.

TcorAA A matrix of epistatic correlations between traits (similar to TcorA). Must be

provided in combination with EcorAA.

Alternatively, corAA can be specified.

EcorAA A matrix of epistatic correlations between environments (similar to EcorA).

Must be provided in combination with TcorAA.

Alternatively, corAA can be specified.

#### **Details**

unstr\_asr\_input can handle separable and non-separable structures between traits and environments.

- For separable structures, provide (1) Tvar & Evar, and (2) TcorA & EcorA.
- For non-separable structures, provide (1) var, and (2) corA.

Note: 'AlphaSimR' can simulate different biological effects (see: SimParam).

- For additive traits use addTraitA().
- For additive + dominance traits use addTraitAD().
- For additive + epistatic traits use addTraitAE().
- For additive + dominance + epistatic traits use addTraitADE().

Check the useVarA argument of these functions when simulating non-additive traits.

#### Value

A list with input parameters for 'AlphaSimR', which are used to simulate correlated genetic values based on an unstructured model for GxE interaction.

```
# Simulate genetic values with 'AlphaSimR' for two additive + dominance traits
# in two environments based on an unstructured model.
# 1. Define the genetic architecture of the simulated traits.
# Mean genetic values and mean dominance degrees.
mean <- c(4.9, 5.4, 235.2, 228.5) # Trait 1 x 2 environments, Trait 2 x 2 environments
meanDD <- c(0.4, 0.4, 0.1, 0.1) # Trait 1 and 2, same value for both environments
# Additive genetic variances and dominance degree variances.
var <- c(0.086, 0.12, 15.1, 8.5) # Trait 1 x 2 environments, Trait 2 x 2 environments
varDD <- 0.2 # Same value for all environment-within-trait combinations
# Additive genetic correlations between the two simulated traits.
TcorA <- matrix(c(</pre>
 1.0, 0.6,
 0.6, 1.0
), ncol = 2)
# Additive genetic correlations between the two simulated environments.
EcorA <- matrix(c(</pre>
 1.0, 0.2,
 0.2, 1.0
), ncol = 2)
# Dominance degree correlations between the four environment-within-trait combinations.
corDD <- diag(4) # Assuming independence</pre>
```

34 unstr\_asr\_output

```
input_asr <- unstr_asr_input(
  ntraits = 2,
  nenvs = 2,
  mean = mean,
  var = var,
  TcorA = TcorA,
  EcorA = EcorA,
  meanDD = meanDD,
  varDD = varDD,
  corDD = corDD
)</pre>
```

unstr\_asr\_output

Simulate genetic values based on an unstructured model for GxE interaction - Simulation with 'AlphaSimR'

### **Description**

Creates a data frame of simulated genetic values in multiple environments for one or more traits based on an unstructured model for genotype-by-environment (GxE) interaction. The wrapper function unstr\_asr\_output requires an 'AlphaSimR' population object generated with unstr\_asr\_input.

### Usage

```
unstr_asr_output(pop, ntraits = 1, nenvs, nreps = 1)
```

# **Arguments**

pop An 'AlphaSimR' population object (Pop-class or HybridPop-class) generated

with unstr\_asr\_input.

ntraits Number of simulated traits specified in unstr\_asr\_input.

nenvs Number of simulated environments specified in unstr\_asr\_input.

nreps A vector defining the number of replicates in each environment. If only one

value is specified, all environments will be assigned the same number.

#### Value

A data frame with columns 'env', genotype 'id', and 'rep', followed by the simulated genetic values for each trait.

# **Examples**

```
# Simulate genetic values with 'AlphaSimR' for two additive + dominance traits
```

# in two environments based on an unstructured model.

# 1. Define the genetic architecture of the simulated traits.

# Mean genetic values and mean dominance degrees.

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```
mean <- c(4.9, 5.4, 235.2, 228.5) # Trait 1 x 2 environments, Trait 2 x 2 environments
meanDD <- c(0.4, 0.4, 0.1, 0.1) # Trait 1 and 2, same value for both environments
# Additive genetic variances and dominance degree variances.
var <- c(0.086, 0.12, 15.1, 8.5) \# Trait 1 x 2 environments, Trait 2 x 2 environments
varDD <- 0.2 # Same value for all environment-within-trait combinations</pre>
# Additive genetic correlations between the two simulated traits.
TcorA <- matrix(c(</pre>
  1.0, 0.6,
  0.6, 1.0
), ncol = 2)
# Additive genetic correlations between the two simulated environments.
EcorA <- matrix(c(</pre>
  1.0, 0.2,
  0.2, 1.0
), ncol = 2)
# Dominance degree correlations between the four environment-within-trait combinations.
corDD <- diag(4) # Assuming independence</pre>
input_asr <- unstr_asr_input(</pre>
 ntraits = 2,
  nenvs = 2,
  mean = mean,
  var = var,
  TcorA = TcorA,
  EcorA = EcorA,
  meanDD = meanDD,
  varDD = varDD,
  corDD = corDD
)
# 2. Use input_asr to simulate genetic values with 'AlphaSimR' based on an
# unstructured model.
library("AlphaSimR")
FOUNDERPOP <- quickHaplo(
  nInd = 10,
  nChr = 1,
  segSites = 20
)
SP <- SimParam$new(FOUNDERPOP)</pre>
SP$addTraitAD(
  nQtlPerChr = 20,
  mean = input_asr$mean,
  var = input_asr$var,
```

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```
corA = input_asr$corA,
 meanDD = input_asr$meanDD,
  varDD = input_asr$varDD,
  corDD = input_asr$corDD,
  useVarA = TRUE
)
# By default, the variances in 'var' represent additive genetic variances.
# When useVarA = FALSE, the values represent total genetic variances.
pop <- newPop(FOUNDERPOP)</pre>
# 3. Create a data frame with simulated genetic values for the two traits in
# the two environments, with two replicates of each genotype.
gv_df <- unstr_asr_output(</pre>
 pop = pop,
 ntraits = 2,
 nenvs = 2,
 nreps = 2
)
```

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